SYNTHESIS AND BIOLOGICAL ACTIVITY OF 7β -(2-AMINO-2-CARBOXY)-ETHYLTHIOACETAMIDO- 7α -METHOXYCEPHALOSPORIN DERIVATIVES

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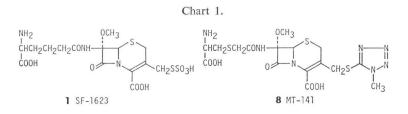
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C-7 and C-3 substituents of a new antibiotic SF-1623 were chemically modified to improve the bioactivity, and substituent effect at C-7 and C-3 was examined on the basis of MIC and ED₅₀ values against selected bacteria. Among a large number of derivatives, MT-141 possessing 2-D-2-amino-2-carboxyethylthioacetamido residue at C-7 and *N*-methyltetrazolylthiomethyl residue at C-3 showed a high order of antibacterial activity *in vitro* and especially *in vivo*.

In the preceding paper¹, we reported the isolation and structural elucidation of a new β -lactam antibiotic SF-1623 (1) belonging to the cephamycin group. Although it was active against some Gramnegative bacteria, the activity of the antibiotic was not high enough for practical application. Successful synthesis of analogs of cephamycin C such as cefoxitin^{2, 8} and cefmetazole^{4, 5} with marked resistance to β -lactamases and broad spectrum activity prompted us to make extensive effort to improve the bioactivity of SF-1623 (1) by chemical modification. Our effort was concentrated on modifying the D-amino-adipoyl group of SF-1623 (1), because alteration of this group has not been extensively studied in the past. The synthetic derivatives were screened not only by *in vitro* evaluation but also *in vivo*, because these properties may not always be in parallel, in particular in β -lactam antibiotics.⁶

In this paper, we report the synthesis of thio analogs of SF-1623 (1) where the central methylene of the α -aminoadipoyl group is replaced by a sulfur atom. The compounds were prepared either by direct modification of 1 or by synthesis from 7β -amino- 7α -methoxycephalosporanic acid. The structure-activity study resulted in the synthesis of MT-141 (8) having potent antibacterial properties.*



Synthesis of Analogs of SF-1623

Semisynthetic cephamycins in this study were prepared by the three general methods $(A \sim C)$

^{*} A part of this paper was presented at the 100th Annual Meeting of the Pharmaceutical Society of Japan, Tokyo, Abstract No. 4J 9–5, 1980.

outlined in Chart 2. Antibiotic SF-1623 (1) was used as a starting material in methods A and B, and 7β -amino- 7α -methoxy-3-cephem compound (XIII) in method C. According to method A, *N*-protected I was converted to the desulfurized exomethylene compound II⁷, which was transformed to V via 3-bromomethyl compound IV³. Application of the acyl exchange method⁹ to V followed by deprotection gave 7β -halogenoacetamido derivative VII, which was easily converted to the desired compound VIII by reaction with an appropriate mercaptan R₄SH.

According to method B, disulfide IX obtained by reaction of N-protected I with sodium p-chlorothiophenolate was reduced with tributylphosphine to afford mercaptan XI, which was converted to V by substitution with a diazoalkane or heterocyclic bromide. Compound V thus obtained could be transformed into VIII by the successive reactions shown in method A.

For the preparation of a variety of semisynthetic cephamycins containing various heterocyclic thio-

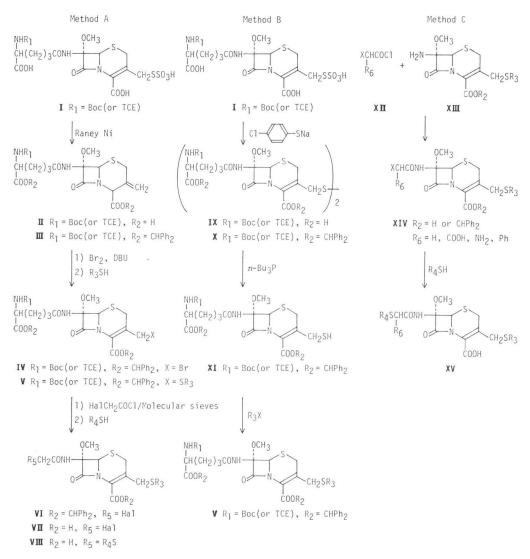


Chart 2.

VOL. XXXVI NO. 3

methyl groups, method C was most suitable. The starting material XIII in this route could be prepared by the known process⁴⁾ or a new process developed in these laboratories¹⁰⁾. Direct acylation or *S*alkylation of halogenoacetyl intermediates (XIV) yielded the final cephamycins (XV). Other synthetic routes that are not comprised in methods $A \sim C$ are described in the experimental section.

Effect of the C-3 Substituents on the Bioactivity

We examined first the effect of C-3 substituents on the antibacterial activity. Six pathogenic bacteria, one Gram-positive and five Gram-negative, were used as test organisms, and the results are listed in Table 1. 7β -Side chain employed in this study was either the 2-D-amino-2-carboxy-ethylthioacetamido group or 2-aminoethylthioacetamido group. The first C-3 substituent examined was the sulfothio group, which is a characteristic moiety of antibiotic SF-1623 (1) and has not been extensively studied. It was found, however, that compounds 3 and 4 having this group showed poor activity except compound 4 against *Proteus vulgaris*.

Replacement of the sulfothio group by an acetoxy group (5 and 6) showed remarkable improvement of the antibacterial activity and spectrum. Replacement of the acetoxy group with azide (10) resulted in further improvement of the activity. However, compound 10 was less active than 8 having a *N*methyltetrazolylthiomethyl group, especially against *Staphylococcus aureus* and *P. vulgaris*. Compounds 13 and 14 possessing neutral heterocycles were also less active than compound 8. Compounds 11 and 12 possessing acidic heterocycles were almost inactive against *S. aureus* but rather active against Gram-negative bacteria. The *N*-dimethylaminoethyltetrazolylthiomethyl group in 15 enhanced the bioactivity of the compound similar to the *N*-methyltetrazolylthiomethyl group in 8 except for *Serratia marcescens*. Compounds 16 and 17 containing the pyridinium moieties were less active against Gramnegative bacteria than 8. Essentially, the same effect of the C-3 substituents was observed when the 7β -side chain was an aminoethylthioacetamido instead of a 2-D-amino-2-carboxyethylthioacetamido group.

Effect of C-7 Substituents on the Bioactivity

Effects of different 7β -side chains on the antibacterial activity *in vitro* are listed in Table 2. All compound had the *N*-methyltetrazolylthiomethyl group at the C-3 substituent. Compound 2 having a D- α -aminoadipoyl side chain was a parent cephamycin to compare with.

Compound 7, in which the 7β -aminoadipoyl group was replaced with a 2-aminoethylthioacetamido group, exhibited improved activity, especially against *S. aureus*, *P. vulgaris* and *S. marcescens* as compared to compound 2. Introduction of a phenyl group in the α -position of the side chain reduced the activity (compound 18).

Notably, replacement of the central methylene of the α -aminoadipoyl group with a sulfur atom as in 8 contributed to greater activity against Gram-negative bacteria, and MICs of 8 were the lowest among the cephamycins studied. The activity against *S. aureus* was also improved. Addition of one more methylene carbon to the C-7 side chain of 8 resulted in a small change in the bioactivity (compound 19). Cyclization of the amino acid function *via* a carbamoyl group caused moderate reduction of the bioactivity (21), but other substitutions such as methoxyimination at α -position (23), carboxylation at α position (24) and dimethylation at γ -position (22) brought about considerable decrease in bioactivity. An amino group in the α -position reduced the activity considerably (25).

We have attempted further modification of the amino acid moiety of compound 8. Introduction of an acyl group, such as *N*-acetyl (26), *N*-carbamoyl (27), *N*-glycyl (28) and *N*-alanyl group (29) caused

THE JOURNAL OF ANTIBIOTICS

		Table 1. Effect o NH/ CHU COU	2 CH ₂ CH ₂ CH ₂ CONH-	OCH3 S	CH ₂ R ₁	,			
					MI	C (µg/ml)			
Compound		R ₁	S. aureus ^{a)}	E. coli ^{b)}	K. pneu- moniae ^{c)}	S. typhi ^d)	P. vulgaris ^{e)}	S. mar- cescens ^f	
1		–SSO ₃ H	200	50	50	12.5	3.13	200	
2		-SUNN CH3	50	6.25	6.25	3.13	1.56	12.5	
			NH2 CHCH2SCH2CONH R2 (CH ₂ R ₁				
	MIC (µg/ml)								
Compound	R_2	R ₁	S. aureus ^{a)}	E. coli ^{b)}			P. vulgaris ^{e)}	S. mar- cescens ^f)	
3	–H	–SSO₃H	200	50	100	50	25	100	
4	-COOH	$-SSO_{3}H$	200	50	50	25	3.13	200	
5	-H	-OAc	12.5	12.5	25	6.25	12.5	25	
6	-COOH	-OAc	100	6.25	6.25	3.13	0.39	50	
7	-H		1.56	3.13	6.25	1.56	0.39	3.13	
8	-COOH	-SUN N	6.25	0.78	1.56	0.39	0.05	1.56	
9	//	-SCH ₃	100	50	100	25	25	100	
10	"	$-N_3$	25	1.56	1.56	0.78	0.20	1.56	
11	"	-s, N, N сн ₂ соон	100	25	6.25	3.13	0.39	12.5	
12	"	S↓_N↓СН₂СООН	100	6.25	3.13	3.13	0.78	50	
13	"	H2 -S H2 CH2	50	3.13	3.13	1.56	0.39	6.25	
14	"	- SH N CH3 - SH SHCH3	25	3.13	1.56	0.78	0.20	12.5	
15	"	-S-CH3 -S-CH2CH2NCH3 -S-CH2CH2NCH3 -S-CH3 -S-CH3 -S-CH3 -S-CH3	12.5	1.56	3.13	1.56	0.78	25	
16	"	-S-CH3	6.25	6.25	12.5	3.13	0.78	3.13	
17	"	-N-CONH2	3.13	12.5	12.5	6.25	3.13	6.25	

Table 1. Effect of C-3 substituents on MIC to six pathogens.

a) Staphylococcus aureus Smith, b) Escherichia coli NIHJ JC-2, c) Klebsiella pneumoniae PCI-602, d) Salmonella typhi O-901-W, e) Proteus vulgaris OX-19, f) Serratia marcescens No. 1.

VOL. XXXVI NO. 3 THE JOURNAL OF ANTIBIOTICS

decrease in the antibacterial activity against all the test organisms. *N*-2-Hydroxybenzyl analog 30, which is a *N*-aralkyl derivative of 8, was 4 times more active than 8 against *S. aureus*, but markedly less active against Gram-negative bacteria. Ester and amido derivatives (31 and 32) of carboxyl function were also $2 \sim 4$ times more active than 8 against *S. aureus*, but they were inferior to 8 against Gram-negative bacteria.

These results indicated that the intact amino acid function of 8 contributed to its high antibacterial activity, especially against Gram-negative bacteria. Compound 8 was coded MT-141.

Effect of 7α -Methoxy Group on the Antibacterial Activity

In a series of compounds having the 7β -2-D-amino-2-carboxyethylthioacetamido group in common, the *in vitro* activities of 7α -hydrogen and 7α -methoxy compounds were compared.

For this purpose, 7α -hydrogen analogs 33, 34 and 35 corresponding to the respective 7α -methoxy congeners 8, 13 and 17 were prepared and their activity examined against 7 pathogens including β -lactamase-producing strains (Table 3). It was found that the 7α -hydrogen compounds had enhanced activity against *S. aureus*, but that they were as active as the 7α -methoxy compound against sensitive Gram-negative bacteria. However, against *P. vulgaris, Proteus morganii* and *S. marcescens*, all of the 7α -methoxy compounds tested were definitely more active than the 7α -hydrogen congeners. An exception was *Pseudomonas aeruginosa*, against which 7α -hydrogen compounds, especially compound 35 possessing pyridinium cation at C-3 were more active than the 7α -methoxy congeners.

Configurational Effect of α -Amino Acid Moiety at C-7

As a result of screening of C-3 and C-7 substituents described above, MT-141 (8) comprising 7β -2-D-2-amino-2-carboxyethylthioacetamido and 3-*N*-methyltetrazolylthiomethyl groups was selected as a promising candidate for further evaluation. However, the 2-amino-2-carboxyethylthioacetamido group contained an asymmetric carbon, and MT-141 (8) existed as the D-isomer. Therefore, it was necessary to compare the activity of the D- and L-isomers, before final selection was made. For this purpose, L-isomers **36**, **37** and **38** corresponding to **8**, **31** and **27** respectively were prepared, and MIC values against six pathogens determined (Table 4).

Among all the compounds tested, MT-141 (8) with D-configuration was the most active. Moreover, it was found that in the three paired compounds listed in Table 4, definite superiority of D-isomer over L-isomer was only observed in the pair 8 and 36, both possessing the intact amino acid moiety. In the other couples, *i.e.* the ethyl ester analogs (31 and 37), and the carbamoyl congeners (27 and 38) D- and L-isomer 31 against some organisms such as *S. aureus* and *S. marcescens*. Thus, acyl and ester derivatization of the α -amino acid moiety caused a loss of the configurational effect that was clearly observed in the original pair, 8 and 36.

As described in the preceding section, substitution of the α -amino acid function of D-congener 8 caused a marked decrease in antibacterial activity. However, this was not observed among the L-isomers. The ester 37 and N-acyl derivative 38 showed almost the same activity as that of the parent 36.

In Vivo Evaluation

In vivo activities of MT-141 (8) and other compounds synthesized in this study were compared on the basis of ED_{50} values which were obtained by using a mouse model intraperitoneally infected with *Escherichia coli* No. 29. The results are summarized in Table 5, together with MIC values. At an

THE JOURNAL OF ANTIBIOTICS

OCH3	c	
R2-CONH	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	NN
0 - N.	CH2S-	N_N_N
	Соон	СН _З

Table 2. Effect of C-7 substituents on MIC to six pathogens.	Table 2.	Effect of	C-7	substituents on	MIC	to	six	pathogens.
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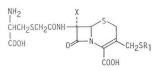
			MIC (µg/ml)							
Compound		R_2	S. aureus	E. coli	K. pneu- moniae	S. typhi	P. vulgaris	S. mar- cescens		
7		NH_{2} CH_{2} CH $_{2}$ SCH $_{2}$ -	1.56	3.13	6.25	1.56	0.39	3.13		
18		$\begin{matrix} \mathbf{NH}_2 & \mathbf{Ph} \\ {}^{I}\mathbf{CH}_2\mathbf{CH}_2\mathbf{SCH} - \end{matrix}$	3.13	6.25	12.5	1.56	12.5	6.25		
8	D	NH2 CHCH2SCH2- COOH NH2	6.25	0.78	1.56	0.39	0.05	1.56		
19	DL	CHCH2CH2SCH2- COOH	6.25	0.78	0.78	0.78	0.20	1.56		
20	D	NH2 CHCH2SCH2CH2- COOH	3.13	3.13	1.56	1.56	1.56	3.13		
21	DL	CONH CHCH ₂ SCH ₂ - NHCO/	3.13	3.13	3.13	0.78	1.56	6.25		
22	D	$\begin{array}{ccc} NH_2 & CH_3 \\ CH_{}C-S-CH_2- \\ COOH & CH_3 \end{array}$	12.5	6.25	6.25	3.13	3.13	6.25		
23	D	NH ₂ ^I CHCH ₂ SCH ₂ C- ^I COOH N-OCH ₃	50	6.25	12.5	6.25	50	12.5		
24	D	NH2 CHCH2SCH- COOH COOH	12.5	6.25	*	3.13	*	50		
25	D	NH2 CHCH2SCH- COOH NH2	100	50	50	50	25	100		
26	D	NHCOCH ₃ CHCH ₂ SCH ₂ – COOH	12.5	3.13	*	0.78	0.39	12.5		
27	D	NHCONH2 CHCH2SCH2- COOH	25	3.13	3.13	1.56	1.56	6.25		
28	D	NHCOCH ₂ NH ₂ CHCH ₂ SCH ₂ - COOH	25	3.13	6.25	1.56	6.25	6.25		
29	D	$\begin{array}{c} CH_3\\ \text{NHCOCHNH}_2\\ \overset{l}{\leftarrow} \text{HCH}_2\text{SCH}_2\text{-}\\ \overset{l}{\leftarrow} \text{OOH}\end{array}$	12.5	6.25	12.5	3.13	1.56	12.5		

			Table 2.	(continued	u)			
30	D	NHCH ₂ –C ₆ H ₄ - <i>o</i> -OH CHCH ₂ SCH ₂ – COOH	1.56	12.5	6.25	3.13	6.25	25
31	D	NH_{2} $\operatorname{CHCH}_{2}\operatorname{SCH}_{2}$ - $\operatorname{COOCH}_{2}\operatorname{CH}_{3}$	3.13	3.13	3.13	1.56	0.78	12.5
32	D	NH2 CHCH2SCH2- CONH2	1.56	3.13	3.13	1.56	3.13	6.25

Table 2. (continued)

* Not determined.

Table 3. Effect of 7α -methoxy group on MIC to eight pathogens.



Com-						MIC	$(\mu g/ml)$			
pound	R1	х	S. aureus	E. coli	K. pneu- moniae	S. typhi	P. vulgaris	P. mor- ganii ^{a)}		P. aeru- ginosa ^{b)}
8	NN	-OCH ₃	6.25	0.78	1.56	0.39	0.05	0.78	1.56	25
33	-SMNN CH3	н	3.13	0.78	0.78	0.78	0.39	50	25	25
13	NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN	$-OCH_3$	50	3.13	3.13	1.56	0.39	1.56	6.25	25
34	-stylo	н	1.56	3.13	1.56	1.56	3.13	50	100	25
17	сн ₃	$-\mathrm{OCH}_3$	3.13	12.5	12.5	6.25	3.13	3.13	6.25	25
35	-N_CONH2	н	1.56	3.13	6.25	6.25	6.25	12.5	12.5	3.13

a) Proteus morganii 1510 (β-Lactamase-producing strain).

b) Pseudomonas aeruginosa MB-3829.

Table 4.	Configurationa	l effect of	α -amino	acid	moiety	on	MIC	to	six	pathogens.
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NHR1	OCH3	N N N
* CHCH2SCH2CONH-	N	N N
I	COOH	I
COOR2 02	COOH	CH3
		0

			Configu-	MIC (µg/ml)							
Compound	R1	R_2	ration (*)	S. aureus	E. coli	K. pneu- moniae	S. typhi	P. vulgaris	S. mar- cescens		
8	н	н	D	6.25	0.78	1.56	0.39	0.05	1.56		
36	H	\mathbf{H}	L	12.5	3.13	6.25	3.13	3.13	6.25		
31	H	Et	D	3.13	3.13	3.13	1.56	0.78	12.5		
37	H	Et	L	1.56	3.13	6.25	1.56	3.13	6.25		
27	CONH_2	\mathbf{H}	D	25	3.13	3.13	1.56	1.56	6.25		
38	CONH_2	Η	L	50	3.13	6.25	3.13	3.13	6.25		

R ₂ CONH CH ₂ R ₁										
Compound		R_2	R1	MIC (µg/ml)	ED₅₀ (mg/mouse)	Geometric standard deviation				
39 *1		NSCH- I COOH	-s LNN CH3	0.20	$\frac{1.0}{(0.57 \sim 1.76)^{*2}}$	2.15				
7		H ₂ NCH ₂ CH ₂ SCH ₂ -	"	1.56	0.50 (0.35 ~0.72)	1.81				
8	D	NH₂ └HCH₂SCH₂- └OOH	"	0.78	0.026 (0.021~0.032)	2.66				
36	L	NH2 CHCH2SCH2- COOH	"	3.13	0.41 (0.234~0.718)	3.57				
31	D	NH2 CHCH2SCH2- COOEt	"	1.56	0.045 (0.021~0.096)	1.10				
32	D	NH2 CHCH2SCH2- CONH2	"	3.13	0.90 (0.60 ~1.35)	1.91				
26	D	NHCOCH ₃ CHCH ₂ SCH ₂ - COOH	"	3.13	0.22 (0.124~0.380)	2.46				
6	D	NH2 CHCH2SCH2- COOH	-OAc	1.56	0.25 (0.143~0.438)	4.66				
10		"	$-N_3$	3.13	0.066 (0.062~0.070)	1.22				
12		"	-S M CH2COOH	6.25	0.15 (0.091~0.248)	3.15				
15		17	-s - s - s - s - s - s - s - s - s - s	1.56	0.052 (0.034~0.08)	1.36				
16		"	-S-(N+-CH3 -S-(N+-CH3) -S-(N+0)	6.25	0.50 (0.33 ~0.75)	3.17				
13		"	-SUN CH3	3.13	0.09 (0.041~0.198)	6.51				
40		H ₂ NCH ₂ CH ₂ SCH ₂ -	"	6.25	>2	—				

Table 5. Comparison of in vitro and in vivo activities to E. coli No. 29.

*1 7α-Hydrogen compound.
*2 Figures in parenthesis indicate 95% confidence limits.

initial stage of this investigation, we have found that compound **39** which was synthesized in the exploratory work, showed a very low MIC value against *E. coli* No. 29, but appeared not as active *in vivo* as expected. This aroused our attention towards the importance of ED_{50} values in evaluating the antibacterial activity of β -lactam antibiotics.

Among the test compounds, MT-141 (8) showed the lowest ED_{50} value, which was far less than that expected from the *in vitro* MIC value. Indeed, 8 was 4 times less active than 39 in MIC value, but 38 times more active in ED_{50} value. Low ED_{50} values were also shown by compounds 31, 15, 10 and 13. All of them contained the same 7β -substituent as 8 or the ethyl ester of it, but the C-3 substituents were varied. Compounds 12 and 16 also exhibited relatively low ED_{50} values in spite of high MIC values.

Configurational and substitutional effect of α -amino acid moiety of the 7 β -side chain was also observed *in vivo*. Compound **36** that is a corresponding L-isomer of **8**, was 16 times less active than D-congener **8** *in vivo*, though the MIC was only 4 times inferior. It was of interest to note that ED₅₀ values of the amido congener **32** and the decarboxyl congener **7** were 20 and 35 times less active than **8** *in vivo*, but 2 and 4 times less active in MIC values. The same was true for compound **40** that was almost inactive *in vivo*, in spite of a MIC of 6.25 μ g/ml.

These results revealed that the 2-D-amino-2-carboxyethylthioacetyl function, which is a sulfur isoster of D-aminoadipoyl moiety in natural SF-1623, played an important role not only for the *in vitro* but also for the *in vivo* activity.

Experimental

Methods

IR spectra were measured in KBr disk using a Hitachi Model 260–10 IR spectrometer. NMR spectra were recorded on a Varian XL-100 spectrometer using TMS as external reference in D_2O unless otherwise stated. The MICs (μ g/ml) of the synthetic cephamycins were determined in two-fold agar dilution method using heart-infusion agar and incubating at 37°C for 20 hours. Inoculum size was 10⁶ colony forming unit (CFU)/ml.

For determination of *in vivo* activity, Slc: ddY female mice of 4 weeks old were used, ten in a group. The animals were inoculated intraperitoneally with 10 LD₅₀ CFU of *E. coli* No. 29 suspended in 2.5% mucin, and were given a single subcutaneous dose of test compound dissolved in distilled water immediately after challenge. The ED₅₀ values (mg/mouse) were determined by the probit method from survival rate of mice after a week.

<u>7β-Chloroacetamido-7α-methoxy-3-(5-methyl-1,3, 4-thiadiazol-2-yl)thiomethyl-3-cephem-4-carbo-</u> xylic Acid (**VIIa**) from SF-1623 (1)

Method A: Two grams of sodium 7β -(5-D-*t*-butoxycarbonylamino-5-carboxyvaleramido)- 7α methoxy-3-sulfothiomethyl-3-cephem-4-carboxylate (I) in 50% aqueous EtOH (50 ml) were hydrogenated at room temperature for 5 hours in the presence of 10 g of RANEY nickel. The catalyst was filtered off and washed with water (50 ml). The filtrate and washings were combined, adjusted to pH 2 with 2 N HCl and extracted with AcOEt (200 ml). The AcOEt extract was washed with water, dried over MgSO₄ and evaporated. The residue was purified by silica gel column chromatography with elution by AcOEt - MeOH (25: 3) to give 1.1 g of 7β -(5-D-*t*-butoxycarbonylamino-5-carboxyvaleramido)- 7α methoxy-3-methylene-3-cephem-4-carboxylic acid (II); IR 1760, 1680 cm⁻¹; NMR (acetone- d_6) δ 1.41 (9H, s, *t*-But), 1.82 (4H, m, CH₂), 2.40 (2H, m, CH₂CO), 3.44 (3H, s, OCH₈), 3.46 (2H, ABq, J=16 Hz, C-2), 4.16 (1H, m, CH), 5.13 (1H, s, C-4), 5.26 (2H, ABq, J=12 Hz, C-3), 5.36 (1H, s, C-6), 6.03 (1H, d, NH), 8.15 (1H, bs, NH).

To a cooled (-78° C) stirred solution of 1,8-diazabicyclo[5.4.0]nndec-7-ene (340 mg) and Br₂ (0.13 ml) in 20 ml of THF were added 615 mg of diphenylmethyl 7 β -(5-D-t-butoxycarbonylamino-5-diphenyl-

methoxycarbonylvaleramido)- 7α -methoxy-3-methylene-3-cephem-4-carboxylate (III) prepared from II in 15 ml THF, and a mixture was kept at -78° C for 20 minutes, and then at 0°C for 20 minutes. After addition of P(OMe)₈ (0.2 ml), the reaction mixture was evaporated and chromatographed over Sephadex LH-20 to give 390 mg of diphenylmethyl 7β -(5-D-*t*-butoxycarbonylamino-5-diphenylmethoxycarbonylvaleramido)- 7α -methoxy-3-bromomethyl-3-cephem-4-carboxylate (IV); IR 1780, 1720, 1670 cm⁻¹; NMR (CDCl₈) δ 1.40 (9H, s, *t*-But), 1.76 (4H, m, CH₂), 2.28 (2H, m, CH₂CO), 3.50 (3H, s, OCH₃), 3.52 (2H, ABq, J=18 Hz, C-2), 4.08 (2H, ABq, J=13 Hz, C-3), 4.40 (1H, m, CH), 5.05 (1H, s, C-6), 6.90 (2H, bs, CH), 7.32 (20H, m, Ph).

To a solution of 350 mg of IV in DMF (5 ml) was added 5-mercapto-2-methyl-1,3,4-thiadiazole (45 mg), and the mixture was stirred at room temperature for 15 hours. The reaction mixture was diluted with AcOEt (70 ml), and washed with water (30 ml×2). The solvent layer was dried over MgSO₄, evaporated, and chromatographed over Sephadex LH-20 to afford diphenylmethyl 7β -(5-D-*t*-butoxycarbonylamino-5-diphenylmethoxycarbonylvaleramido)- 7α -methoxy-3-(5-methyl-1,3,4-thiadiazol-2-yl)thiomethyl-3-cephem-4-carboxylate (V); IR 1780, 1710, 1680 cm⁻¹; NMR (CDCl₃) δ 1.40 (9H, s, *t*-But), 1.75 (4H, m, CH₂), 2.20 (2H, m, CH₂CO), 3.49 (3H, s, OCH₃), 2.73 (3H, s, CH₃), 3.56 (2H, bs, C-2), 4.15 (2H, ABq, J=13 Hz, C-3), 4.43 (1H, m, CH), 5.02 (1H, s, C-6), 6.88 (1H, s, CH), 6.90 (1H, s, CH), 7.32 (20H, m, Ph).

Compound V (190 mg) was dissolved in 6 ml of dichloromethane, and to this solution were added Molecular Sieves 4A (1.9 g) and chloroacetyl chloride (100 mg), and a mixture was stirred at room temperature for 48 hours. The reaction mixture was filtered off and the filtrate was washed with 10% aqueous NaHCO₃ and water. The solvent layer was dried over MgSO₄, and evaporated to give 210 mg of diacyl compound, which was subsequently treated with a mixture of CF₃COOH (1.5 ml) and anisole (3 ml) at 0°C for 50 minutes to afford 7 β -chloroacetamido-7 α -methoxy-3-(5-methyl-1,3,4-thiadiazol-2yl)thiomethyl-3-cephem-4-carboxylic acid (VIIa); IR 1775, 1680, 1610 cm⁻¹; NMR (acetone- d_6) δ 2.76 (3H, s, CH₃), 3.54 (3H, s, OCH₃), 3.70 (2H, ABq, J=18 Hz, C-2), 4.26 (2H, s, CH₂CO), 4.45 (2H, ABq, J=14 Hz, C-3), 5.12 (1H, s, C-6).

 7β -Bromoacetamido- 7α -methoxy-3-methylthiomethyl-3-cephem-4-carboxylic Acid (VIIb) from SF-1623 (1)

Method B: A mixture of 3.0 g of 7β -(5-D-*t*-butoxycarbonylamino-5-carboxyvaleramido)- 7α -methoxy-3-sulfothiomethyl-3-cephem-4-carboxylic acid (I) and sodium *p*-chlorothiophenolate (0.8 g) in water (50 ml) was stirred at room temperature for 2.5 hours.

When the reaction was over, the mixture was washed with AcOEt (30ml), and the aqueous layer was adjusted to pH 2 with 1 N HCl, and extracted with AcOEt (50 ml×2). The solvent layer was washed with water, dried over MgSO₄ and evaporated to give 2.25 g of 1,1'-bis [7 β -(5-D-*t*-butoxycarbonylamino-5-carboxyvaleramido)-7 α -methoxy-4-carboxy-3-cephem-3-yl]dimethyl disulfide (IX); IR 1760, 1705, 1620 cm⁻¹; NMR (acetone- d_0) δ 1.40 (9H, s, *t*-But), 1.90 (4H, m, CH₂), 2.45 (2H, m, CH₂CO), 3.51 (3H, s, OCH₃), 3.56 (2H, ABq, J=18 Hz, C-2), 3.97 (2H, ABq, J=13 Hz, C-3), 4.25 (1H, m, CH), 5.11 (1H, s, C-6), 7.05 (1H, d, NH), 8.17 (1H, bs, CONH).

To a solution of 850 mg of 1,1'-bis[7 β -(5-D-*t*-butoxycarbonylamino-5-diphenylmethoxycarbonylvaleramido)-7 α -methoxy-4-diphenylmethoxycarbonyl-3-cephem-3-yl]dimethyl disulfide (X) prepared from IX in 40% aqueous dioxane (25 ml) was added *n*-tributyl phosphine (0.25 ml), and the mixture was stirred at 40°C for 45 minutes. The reaction mixture was evaporated, diluted with water (40 ml) and extracted with AcOEt (100 ml). The solvent layer was dried over MgSO₄, evaporated and chromatographed over Sephadex LH-20 with elution by AcOEt to give 720 mg of diphenylmethyl 7 β -(5-D-*t*butoxycarbonylamino-5-diphenylmethoxycarbonylvaleramido)-7 α -methoxy-3-thiomethyl-3 -cephem-4carboxylate (XI); IR 2550, 1770, 1720, 1685 cm⁻¹; NMR (CDCl₃) δ 1.40 (9H, s, *t*-But), 1.73 (4H, m, CH₂), 1.88 (1H, dd, SH), 2.20 (2H, m, CH₂CO), 3.47 (2H, bs, C-2), 2.92 & 3.72 (2H, each dd, C-3), 3.52 (3H, s, OCH₃), 4.42 (1H, m, CH), 5.04 (1H, s, C-6), 6.90 (1H, s, CH), 6.92 (1H, s, CH), 7.32 (20H, m, Ph).

To a solution of XI (640 mg) in AcOEt (30 ml) was added an ether solution of diazomethane at 0°C. When the reaction was completed, the mixture was evaporated and chromatographed over silica gel with benzene - AcOEt (5: 1) as a developing solvent to give 530 mg of diphenylmethyl 7β -(5-D-*t*-butoxycarbonylvaleramido) - 7α -methoxy- 3 -methylthiomethyl- 3 -cephem-4 - Application of the known diacyl exchange reaction to V (440 mg) as described above using bromoacetyl chloride afforded 125 mg of 7β -bromoacetamido- 7α -methoxy-3-methylthiomethyl-3-cephem-4carboxylic acid (**VIIb**); IR 1760, 1670, 1600 cm⁻¹; NMR (acetone- d_e) δ 2.0 (3H, s, CH₃), 3.44 (2H, ABq, J=18 Hz, C-2), 3.52 (3H, s, OCH₃), 3.61 (2H, ABq, J=13 Hz, C-3), 4.09 (2H, s, CH₂CO), 5.16 (1H, s, C-6).

 7β -(2-D-Amino-2-carboxyethylthioacetamido)- 7α -methoxy-3-(1-methyl-1*H*-tetrazol-5-yl)thiomethyl-3-cephem-4-carboxylic Acid (8)

Method C: A mixture of 7β -bromoacetamido- 7α -methoxy-3-(1-methyl-1*H*-tetrazol-5-yl)thiomethyl-3-cephem-4-carboxylic acid (XIV) (920 mg) and D-cysteine hydrochloride (470 mg) in water (15 ml) was adjusted to pH 6.8 with aqueous NaHCO₃, and stirred under ice-cooling for 2 hours. The reaction mixture was chromatographed on a column of Diaion HP-20 with elution by water to give 1.1 g of sodium salt of **8** which was crystallized from H₂O - EtOH; mp 90~91°C; *Anal.* Calcd. for C₁₆-H₂₀N₇O₇S₈Na·7H₂O: C 28.78, H 5.13, N 14.68, Found: C 28.84, H 4.96, N 14.21; IR 1755, 1670, 1610 cm⁻¹; NMR (D₂O) δ 3.22 (2H, m, CH₂S), 3.50 (2H, s, SCH₂CO), 3.55 (3H, s, OCH₃), 3.59 (2H, ABq, *J*=18 Hz, C-2), 3.97 (1H, m, CH), 4.04 (3H, s, N-CH₃), 4.16 (2H, ABq, *J*=13 Hz, C-3), 5.15 (1H, s, C-6).

Similar reaction as described above, using DL-homocysteine, DL-cysteinehydantoin, D-penicillamine, D-cysteine ethyl ester, D-cysteine amide and L-cysteine in place of D-cysteine afforded the respective products 19, 21, 22, 31, 32, and 36.

 $\frac{7\beta - (2-D-Amino - 2 - carboxyethylthioacetamido) - 7\alpha - methoxy - 3 - acetoxymethyl - 3 - cephem - 4 - carbo$ xylic Acid (6)

Treatment of 7β -bromoacetamido- 7α -methoxycephalosporin (425 mg) with D-cysteine (200 mg) by a similar procedure as described for the preparation of **8** gave 320 mg of sodium salt of **6**; IR 1760, 1670, 1605 cm⁻¹; NMR (D₂O) δ 2.10 (3H, s, OAc), 3.20 (2H, m, CH₂S), 3.50 (2H, ABq, J=18 Hz, C-2), 3.51 (2H, s, SCH₂CO), 3.57 (3H, s, OCH₃), 3.98 (1H, m, CH), 4.78 (2H, ABq, J=13 Hz, C-3), 5.19 (1H, s, C-6).

 7β -(2-D-Amino-2-carboxyethylthioacetamido)- 7α -methoxy-3-(heterocyclicthiomethyl or pyridiniummethyl)-3-cephem-4-carboxylic Acid (11, 12, 15, 16 and 17) by Substitution Reaction of 6

General Procedure: A mixture of 485 mg (1 mmole) of sodium salt of **6** and heterocyclic thiol or substituted pyridine (each 2 mmole) was dissolved in 15 ml of 0.1 M phosphate buffer (pH 6.8), and stirred at $60 \sim 65^{\circ}$ C for 6 hours under nitrogen atmosphere. When the reaction was over, the mixture was chromatographed on Diaion HP-20 column with elution by water to give desired compounds. Spectral data of some typical compounds were as follows.

 7β -(2-D-Amino- 2-carboxyethylthioacetamido)- 7α -methoxy- 3-(1-carboxymethyl-1*H*-tetrazol- 5-yl)-thiomethyl-3-cephem-4-carboxylic Acid (11)

IR 1755, 1610 cm⁻¹; NMR (D₂O) δ 3.16 (2H, m, CH₂S), 3.44 (2H, s, SCH₂CO), 3.55 (3H, s, OMe), 3.59 (2H, ABq, J=18 Hz, C-2), 3.92 (1H q, CH), 4.21 (2H, ABq, J=13 Hz, C-3), 5.00 (2H, s, N-CH₂), 5.15 (1H, s, C-6).

 7β -(2-D-Amino- 2 -carboxyethylthioacetamido)- 7α -methoxy- 3 -(4-carbamoylpyridinium)methyl- 3 - cephem-4-carboxylic Acid (17)

IR 1760, 1670, 1620 cm⁻¹; NMR (D₂O) δ 3.20 (2H, m, CH₂S), 3.47 (2H, s, SCH₂CO), 3.55 (3H, s, OCH₃), 3.57 (2H, ABq, J=18 Hz, C-2), 3.95 (1H, m, CH), 5.20 (1H, s, C-6), 5.48 (2H, ABq, J=13 Hz, C-3), 8.37 & 9.11 (4H, each d, J=6 Hz, pyridinium)

 7β -[3-(2-D-Amino- 2 -carboxyethylthio)- 2 -methoxyimino]propionamido- 7α -methoxy- 3 -(1-methyl-1*H*-tetrazol-5-yl)thiomethyl-3-cephem-4-carboxylic Acid (23)

To an ice-cooled solution of 525 mg of diphenylmethyl 7 β -amino-7 α -methoxy-3-(1-methyl-1H-

tetrazol-5-yl)thiomethyl-3-cephem-4-carboxylate (XIII) and N,N-dimethylaniline (0.15 ml) in CH₂Cl₂ (25 ml) was added dropwise under stirring 3-bromo-2-methoxyiminopropionyl chloride (215 mg) in CH₂Cl₂ (5 ml).

After reaction at 10°C for 3 hours, the mixture was washed sequentially with 0.2 N HCl, aqueous 5% NaHCO₃ and water. The solvent layer was dried over MgSO₄ and evaporated to give 630 mg of diphenylmethyl 7 β -(3-bromo-2-methoxyimino)propionamido-7 α -methoxy-3-(1-methyl-1*H*-tetrazol-5-yl)-thiomethyl-3-cephem-4-carboxylate; IR 1760, 1720, 1670 cm⁻¹; NMR (CDCl₃) δ 3.54 (3H, s, OMe), 3.60 (2H, ABq, J=18 Hz, C-2), 4.01 (3H, s, N-CH₃), 4.04 (3H, s, OCH₃), 4.22 (2H, s, BrCH₂C), 4.25 (2H, ABq, J=13 Hz, C-3), 5.09 (1H, s, C-6), 6.90 (1H, s, CH), 7.30 (10H, m, Ph).

Removal of diphenylmethyl group from this ester (350 mg) by treatment with CF₃COOH and anisole, followed by reaction with D-cysteine gave 210 mg of 23; IR 1760, 1660, 1620 cm⁻¹; NMR (D₂O) δ 3.10 (2H, m, CH₂S), 3.55 (3H, s, OCH₃), 3.60 (2H, ABq, *J*=18 Hz, C-2), 3.62 (2H, bs, SCH₂), 3.95 (1H, m, CH), 4.01 (3H, s, N-CH₃), 4.04 (3H, s, OCH₃), 4.18 (2H, ABq, *J*=13 Hz, C-3), 5.14 (1H, s, C-6).

Compounds 18 and 24 were prepared analogously using α -bromophenylacetyl chloride and half acid chloride of bromomalonic acid, respectively.

 7β -(2-D-Carbamoylamino-2-carboxyethylthioacetamido)- 7α -methoxy-3-(1-methyl-1*H*-tetrazol-5-yl)thiomethyl-3-cephem-4-carboxylic Acid (27)

A mixture of sodium salt of 8 (300 mg) and 150 mg of potassium cyanate in water (20 ml) was stirred at room temperature for 24 hours. The reaction mixture was subjected to a column chromatography of Diaion HP-20 developing with water to give sodium salt of 27 (180 mg); IR 1770, 1660, 1610 cm⁻¹; NMR (D₂O) δ 3.07 (2H, m, CH₂S), 3.47 (2H, s, SCH₂CO), 3.56 (3H, s, OCH₃), 3.60 (2H, ABq, *J*=18 Hz, C-2), 4.05 (3H, s, N-CH₃), 4.17 (2H, ABq, 14 Hz, *J*=13 Hz, C-3), 4.22 (1H, m, CH), 5.14 (1H, s, C-6).

 7β -[2-D- (Aminomethylcarbonyl)amino - 2 - carboxyethylthioacetamido] - 7α - methoxy- 3 -(1-methyl-1*H*-tetrazol-5-yl)thiomethyl-3-cephem-4-carboxylic Acid (28)

To an ice-salt cooled solution of *N*-*t*-butoxycarbonylglycine (175 mg) in THF (7 ml) were added triethylamine (0.14 ml) and ethyl chloroformate (0.1 ml), and the mixture was stirred at -20° C for 40 minutes. After addition of sodium salt of **8** (405 mg) dissolved in 50% aqueous THF (8 ml), the mixture was stirred at -20° C for 30 minutes, and then at $0 \sim 5^{\circ}$ C for 2 hours. When the reaction was over, the solution was diluted with water (20 ml), layered over with AcOEt (70 ml), brought to pH 2 with 1 N HCl and shaken under cooling. The AcOEt extract was washed with water, and evaporated to give 496 mg of *N*-*t*-butoxycarbonylglycyl derivative of **8** (**28a**); IR 1760, 1670, 1605 cm⁻¹; NMR (acetone- $d_{\rm g}$) δ 1.45 (9H, s, *t*-But), 3.19 (2H, m, CH₂S), 3.48 (2H, s, SCH₂CO), 3.55 (3H, s, OCH₃), 3.70 (2H, ABq, *J*=18 Hz, C-2), 3.82 (2H, bs, CH₂NH), 4.03 (3H, s, N-CH₃), 4.45 (2H, ABq, *J*=13 Hz, C-3), 4.80 (1H, m, CH), 5.11 (1H, s, C-6).

A cooled solution of **28**a (450 mg) in CHCl₃ (2 ml) was treated with CF₃COOH (4 ml) at 0°C for 30 minutes, and evaporated. The residue was washed with *n*-hexane, dissolved in small amount of water and adjusted to pH 6 with 1 N NaOH. Chromatographic purification over Diaion HP-20 afforded 150 mg of sodium salt of **28**; IR 1760, 1670, 1610 cm⁻¹; NMR (D₂O) δ 3.10 (2H, m, CH₂S), 3.43 (2H, s, SCH₂), 3.53 (3H, s, OCH₃), 3.57 (2H, ABq, J=18 Hz, C-2), 3.87 (2H, bs, CH₂NH), 4.03 (3H, s, N-CH₃), 4.16 (2H, ABq, J=13 Hz, C-2), 4.46 (1H, m, CH), 5.12 (1H, s, C-6).

 7β -[2-D-(2-Hydroxybenzylamino) - 2 - carboxyethylthioacetamido] - 7α - methoxy- 3 - (1-methyl- 1*H*-tetrazol-5-yl)thiomethyl-3-cephem-4-carboxylic Acid (**30**)

To a mixture of 8 (295 mg) and salicylaldehyde (70 mg) in 50% aqueous MeOH was added NaBH₄ (50 mg), and a solution was stirred at $5 \sim 10^{\circ}$ C for 3 hours. The reaction mixture was evaporated and chromatographed over Diaion HP-20 with elution by 20% aqueous EtOH to give 180 mg of **30**; IR 1755, 1610 cm⁻¹; NMR (D₂O-acetone- d_0) δ 3.20 (2H, m, CH₂S), 3.45 (2H, bs, SCH₂), 3.55 (3H, s, OCH₃), 3.61 (2H, ABq, J=18 Hz, C-2), 3.81 (2H, bs, NHCH₂), 4.03 (3H, s, N-CH₃), 4.20 (1H, m, CH), 4.26 (2H, ABq, J=13 Hz, C-3), 5.12 (1H, s, C-6), 6.91 & 7.28 (4H, each m, Ph).

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